

CALCIUM PHOSPHATE NANOSYSTEMS AS VEHICLES FOR CONTROLLED DRUG DELIVERY

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Systemic drug administration to bone tissue has several drawbacks as low drug absorption, which leads to supplying high concentrations that damage other tissues. Calcium phosphate (CaP)-based nanosystems can be used for drug transport and delivery to calcium-containing surfaces, as bone mineral matrix, due to its high affinity for hydroxyapatite. With this basis we developed nanospheres based on liposomes of 1,2-Dioleoyl-sn-glycero-3-phosphate (DOPA), coated with a CaP amorphous layer (CaPLi) which provides them with stability, stiffness and affinity in the biological media. The antibiotic Levofloxacin (Lx) was used as a model drug for encapsulation (CaPLiLx), tracking and release studies, which allowed monitoring by fluorescence spectroscopic methods.

The CaPLi were characterized by TEM, SEM, AFM and HR-STEM microscopies, among other techniques.

The CaPLiLx charge release was studied in the PBS and acetate buffers and in simulated body fluid (SBF), following the concentration of Lx by fluorescence. A faster release was evidenced in SBF, where Ca^{2+} and Mg^{2+} ions are present, and we proposed that the divalent cations can complex the surface of the CaP nanolayer, stimulating the release of the drug. The interaction with surfaces (Ca^{2+} -modified mica surface, hydroxyapatite nanoparticle (Ap) films on glass, and Ap modified 45S5[®] bioactive glassbased scaffolds) was evaluated, showing the preferential accumulation of the CaPLi on the calcium-enriched ones, the disruption of the inorganic structure and release of molecules contained in the nanospheres.

Furthermore, biological studies were performed. Bacterial susceptibility and time killing assays with *Staphylococcus aureus* suspensions demonstrated the bactericidal potential of the nanoshells containing Lx (1.0 $\mu\text{g}/\text{ml}$, corresponding to 2x MIC). Moreover, cell viability was analyzed in MC3T3 cell cultures exposed to nanospheres, concluding that they are not cytotoxic under the performed conditions (CaP 200 $\mu\text{gCa}/\text{ml}$ and CaPLiLx 100 μM Lx dilutions up 20% in cell culture). Finally, zebrafish model will be used to assess the *in vivo* biocompatibility and localization aiming to develop stable and functional vehicles that contribute to the optimization of the treatment of bone diseases.